

Disaccharides to Inositol Saccharides: Synthesis of α -D-Galactopyranosyl-D-*myo*-inositol Derivatives from a Methyl 4-O-(α -D-Galactopyranosyl)- α -D-glucopyranoside Derivative

Hari Babu Mereyala* and Sreenivasulu Guntha

Indian Institute of Chemical Technology, Hyderabad 500 007, India

The disaccharide derivative **3** has been transformed into the galactopyranosylcyclohexanone derivative **5** by means of Ferrier carbocycle reaction of the enosaccharide **4**. Compound **5** was converted into the enone **6** and then into the allylic alcohol **7**. Oxidation of compound **7** and the acetyl derivative **10** by OsO₄ gave the galactopyranosyl-D-*myo*-inositol derivatives **8** and **9**, and **11** and **12**, respectively, in good yields.

Glycosylphosphatidylinositol (GPI) membrane anchors are ubiquitous throughout eukaryotic evolution and are found attached to a wide variety of cell-surface glycoproteins.¹ GPIs have been implicated in a second messenger mechanism for signal transduction of insulin.² Among mycobacterial lipids,³ phosphatidyl-*myo*-inositol mannosides (PIMs) have been shown to be antigenic and to elicit protective immunity against experimental tuberculosis when injected alone or with carrier protein.⁴ Inositolsaccharide structural units are also common to several important cyclitol antibiotics.⁵ Plants contain 1L-1-(*O*-D-galactopyranosyl)-*myo*-inositol in which *myo*-inositol functions as a galactosyl transfer co-factor.⁶

In spite of their interesting biological functions very few reports have appeared on their methods to synthesis. They have so far been synthesized basically in two key steps, namely (i) resolution of the *meso*-*myo*-inositol to the D and L forms followed by (ii) stereoselective glycosylation.⁷⁻¹² Thus, the phosphatidyl-*myo*-inositol mannoside residue of *Trypanosoma Brucei* was earlier synthesized by glycosylation of the appropriately protected D-*myo*-inositol alcohol derivative (acceptor) with per-*O*-benzylated pentenyl mannopyranoside (donor) by use of *N*-iodosuccinimide as a promoter.^{7,8} Likewise glycosylation of the D- and L-*myo*-inositol alcohol derivative with a galactosyl chloride derivative in the presence of silver triflate as a promoter resulted in the synthesis of galactosyl D- and L-*myo*-inositol derivatives.⁹ The enantiomeric D- and L-*myo*-inositol alcohol derivatives themselves required for glycosylation were obtained by resolution of DL-*myo*-inositol as its L-(+)-mandelic acid¹⁰ or camphanic acid¹¹ ester or as monoglycosides.^{8,12}

We have developed a highly elegant route where inositol-saccharides were synthesized starting from disaccharides, thereby avoiding cumbersome resolution and/or glycosylation steps (Scheme 1). Feasibility of this scheme is dependent upon the ability to transform one of the pyranose sugars (reducing sugar) of the disaccharide to a carbocycle without damaging the interglycosidic linkage. The easy availability of various disaccharides either in natural abundance or by proven synthetic methods makes this approach highly attractive.

Thus, coupling of 2-pyridyl 1-thiogalactopyranosyl donor **1** with the suitably protected methyl α -D-glucopyranoside **2** by the iodomethane activation procedure¹³ gave the α -linked disaccharide derivative **3** in 86% yield (Scheme 2).

Formation of compound **3** was evident from the ¹H NMR spectrum by the appearance of a signal for 1-H at δ 5.61 with a coupling constant of 3.95 Hz ($J_{1,2}$) and also from the ¹³C NMR spectrum (see Experimental section). Compound **3** was converted into 5-enodisaccharide **4** in a one-pot reaction with NaI

in dimethyl sulfoxide (DMSO) followed by treatment with (DBU) 1,8-diazabicyclo[5.4.0.]undec-7-ene.¹⁴ Compound **4** was characterised by the appearance of C-5 and C-6 signals at δ_c 153.4 and 97.7, respectively, in the ¹³C NMR spectrum. Compound **4** was dissolved in acetone-water (2:1 ratio) and treated with a catalytic amount of Hg(OCOCF₃) at room temperature for 12 h to obtain the Ferrier cyclisation¹⁵ product **5** as an isomeric mixture (10/1 α -alcohol/ β -alcohol) in 92% yield. Appearance of signals for 6-H_{ax} and 6-H_{eq} at δ 2.18 and δ 2.60, respectively, each as a doublet of doublets in the ¹H NMR spectrum, and the carbonyl absorption at 1710 cm⁻¹ in the IR spectrum confirmed the formation of compound **5**. It was as such treated with methanesulfonyl chloride in pyridine to give the enone **6** in 81% yield.¹⁶ Appearance of signals for 3-H and 2-H at δ 5.98 (dd, $J_{2,3}$ 2.25 Hz, $J_{2,6}$ 2.9 Hz) and δ 6.79 (dd, $J_{4,3}$ 2.25 Hz) in the ¹H NMR spectrum of compound **6**, and the presence of an α,β -unsaturated enone absorption at 1660 cm⁻¹ in the IR spectrum, confirmed the formation of compound **6**. Stereoselective reduction of compound **6** was achieved by reaction with CeCl₃·7H₂O in methanol at -78 °C for 2 h followed by addition of NaBH₄ to obtain compound **7** in 87% yield.¹⁷ The alcohol **7** was characterised from the appearance of a signal for 5- and 4-H at δ 5.58 as a AB-type quartet with a coupling of 10 Hz in the ¹H NMR spectrum. Oxidation of ene **7** with OsO₄¹⁸ in acetone/water gave the diastereoisomeric α -D-galactopyranosyl-D-*myo*-inositol derivatives **8** and **9** in the ratio 2:1 in 91% yield. The diastereoisomeric ratio improved to 5.6:1 when the allylic hydroxy group of compound **7** was substituted with a small electron-withdrawing (acetyl) group¹⁹ and then oxidised with OsO₄. Thus, compound **7** was acetylated to give acetate **10** in quantitative yield, and this reacted with OsO₄ to give α -D-galactopyranosyl-D-*myo*-inositol derivatives **11** and **12** (92% yield) as an inseparable mixture. Compounds **11** and **12** were converted into the dibenzoyl derivatives **13** and **14**, respectively, for characterisation by ¹H NMR spectroscopy. 3-H and 2-H in compound **13** appear at δ 5.18 and δ 5.82, respectively, with coupling constants $J_{3,4}$ 9.48, $J_{2,3}$ 2.60 and $J_{1,2}$ 2.68 Hz, whereas in compound **14** 2-H and 3-H appear at δ 6.12 and δ 5.46 with a couplings $J_{1,2} = J_{2,3} = 2.72$ and $J_{3,4}$ 9.59 Hz.

This protocol in principle makes the synthesis of any inositol-saccharide highly practical by the appropriate choice of disaccharide.

Experimental

¹H NMR spectra were measured with a Varian Gemini (200 MHz) spectrometer, with tetramethylsilane as internal standard

(OMe), 67.9, 68.7, 69.7, 70.2, 72.7, 73.3 (2C), 73.5, 73.8, 74.3, 74.5, 74.6, 75.3, 78.9, 79.8 and 81.2 (C-2/6, C-2'/6', and 6 × OCH₂Ph), 97.4 and 97.6 (C-1, -1') and 125.0–140.0 (aromatic).

Methyl 2,3-Di-O-benzyl-6-deoxy-4-O-(2,3,4,6-tetra-O-benzyl- α -D-galactopyranosyl)- α -D-xylo-hex-5-enopyranoside 4.—A mixture of compound 3 (2.09 g, 1.9 mmol), Bu₄Ni (1.42 g, .95 mmol), sodium iodide (1.42 g, 9.5 mmol), and powdered molecular sieves (4 Å; 500 mg) in dry DMSO (20 cm³) was heated to 80 °C. After 2 h, DBU (0.34 cm³, 2.28 mmol) was added and the mixture was heated for another 2 h. When TLC indicated completion of the reaction the mixture was filtered on Celite, washed with ethyl acetate (5 cm³), diluted with water (100 cm³), and extracted into ethyl acetate (100 cm³). The extract was washed with water, dried, and concentrated to obtain *title compound 4* (1.0 g, 60%) as a syrup (Found: C, 75.1; H, 6.6. C₅₄H₅₈O₁₀ requires C, 75.13; H, 6.67%); [α]_D +27 (c 1.0, CHCl₃); δ_{H} (200 MHz; CDCl₃) 3.39 (3 H, s, OMe), 3.45–4.20 and 4.43–5.04 (24 H, m, 1–4-H, 6-H₂, 2'–6'-H and 6 × CH₂Ph), 5.75 (1 H, d, J_{1,2}: 3.51, 1'-H) and 7.15–7.60 (30 H, ArH); δ_{C} (50 MHz; CDCl₃) 55.4 (OMe), 69.5, 70.1, 72.4, 73.1, 73.3, 73.8, 74.3, 74.7, 74.8, 75.1, 75.4, 75.9, 79.0 and 81.5 (C-2/4, C-2'/6' and 6 × CH₂Ph), 97.4 and 98.9 (C-1, -1'), 97.7 (C-6), 125.0–140.0 (aromatic) and 153.4 (C-5).

3,4-Dibenzyl-5-hydroxy-2-(2,3,4,6-tetra-O-benzyl- α -D-galactopyranosyloxy)cyclohexanone 5.—A catalytic amount of mercury(II) trifluoroacetate (10 mg) was added to a solution of the enol ether 4 (2.8 g, 3.19 mmol) in acetone–water (90 cm³; 2:1) and the mixture was left at room temperature for 12 h. It was then concentrated to 30 cm³, diluted with water (100 cm³), and extracted into ethyl acetate (100 cm³). The extract was washed successively with aq. KI (10%), aq. 'hypo' (sodium thiosulfate) (20%) and saturated aq. NaHCO₃, dried, and concentrated to obtain *title compound 5* (2.53 g, 92%; α : β 10:1) as a syrup (Found: C, 74.9; H, 6.5. C₅₄H₅₆O₁₀ requires C, 74.95; H, 6.55%); [α]_D +54 (c 1.0, CHCl₃); ν_{max} (CHCl₃)/cm⁻¹ 1710 (carbonyl); δ_{H} (200 MHz; CDCl₃) 2.18 (1 H, dd, J_{6,6\text{gem}}} 14.24, J_{5,6a} 3.56, 6-H_{ax}), 2.60 (1 H, dd, J_{5,6e} 3.92, 6-H_{eq}), 3.35–5.05 (23 H, m, 2–5-H, 2'–6'-H, OH and 6 × CH₂Ph), 5.35 (1 H, d, J_{1,2}: 3.2, 1'-H) and 7.12–7.50 (30 H, ArH); δ_{C} (50 MHz; CDCl₃) 42.6 (C-6), 66.3, 69.5, 69.9, 72.7, 72.9, 73.0, 73.2, 73.5, 74.6, 74.8, 75.5, 79.4, 80.7, 81.8 and 82.1 (C-2/5, C-2'/6' and 6 × OCH₂Ph), 98.7 (C-1'), 124.0–140.0 (aromatic) and 202.6 (CO).

4,5-Dibenzyl-6-(2,3,4,6-tetra-O-benzyl- α -D-galactopyranosyloxy)cyclohex-2-enone 6.—To a stirred solution of compound 5 (1 g, 1.17 mmol) and 4 (dimethylamino)pyridine (DMAP) (5 mg) in dry pyridine (35 cm³) at 0 °C was added dropwise methanesulfonyl chloride (0.1 cm³, 1.75 mmol). It was brought to room temperature, stirred for 2 h, then diluted with dichloromethane. The organic phase was washed with water, dried, and evaporated to obtain *title compound 6* (0.79 g, 81%) as a syrup (Found: C, 76.5; H, 6.4. C₅₄H₅₄O₉ requires C, 76.54; H, 6.46%); [α]_D +93 (c 1.0, CHCl₃); δ_{H} (200 MHz; CDCl₃) 3.40–5.05 (21 H, m, 4–6-H, 2'–6'-H and 6 × CH₂Ph), 5.50 (1 H, d, J_{1,2}: 3.6, 1'-H), 5.98 (1 H, dd, J_{2,3} 9.85, J_{2,6} 2.9, 2-H), 6.79 (1 H, dd, J_{4,3} 2.25, 3-H) and 7.00–7.45 (30 H, ArH); δ_{C} (50 MHz; CDCl₃) 70.2, 73.2, 73.5, 74.8, 75.0, 75.2, 75.3, 76.0, 76.1, 77.0, 78.2, 78.4, 79.0 and 85.4 (C-4/6, C-2'/6' and 6 × CH₂Ph), 98.7 (C-1'), 128.1 (C-2), 126.0–139.0 (aromatic), 147.5 (C-3) and 195.1 (CO).

1D-(1,3/2,4)-1,2-Di-O-benzyl-3-O-(2,3,4,6-tetra-O-benzyl- α -D-galactopyranosyl)cyclohex-5-ene-1,2,3,4-tetraol 7.—To a stirred solution of compound 6 (0.9 g, 1.06 mmol) in methanol (30 cm³) at –78 °C was added CeCl₃·7H₂O (0.35 g, 0.96 mmol)

and the mixture was stirred for a further 20 min. NaBH₄ (0.23 g, 6.17 mmol) was added and 2 h later the mixture was quenched by addition of acetic acid (0.2 cm³), concentrated to a syrup, and subjected to column chromatography [SiO₂; hexane–ethyl acetate (2:1)] to obtain *title compound 7* (0.78 g, 87%) as a syrup (Found: C, 76.3; H, 6.6. C₅₄H₅₆O₉ requires C, 76.36; H, 6.68%); [α]_D +39 (c 1.0, CHCl₃); δ_{H} (200 MHz; CDCl₃) 3.25–5.00 (21 H, m, 1–2-H, 6-H, 2'–6'-H, OH and 5.5 × CH₂Ph), 5.12 (1 H, d, J_{1,2}: 3.4, 1'-H), 5.15 (1 H, d, CH₂Ph), 5.58 (2 H, AB-type quartet, J_{4,5} 10.0, 5- and 4-H) and 7.10–7.45 (30 H, ArH); δ_{C} (50 MHz; CDCl₃) 69.6, 70.7, 71.5, 72.4, 73.2, 73.5, 75.0, 75.4, 76.3, 76.9, 77.6, 78.1, 79.4, 82.1 and 87.6 (C-1/3, C-6, C-2'/6' and 6 × CH₂Ph), 99.2 (C-1'), 126.0–140.0 (aromatic), 129.4 and 138.3 (C-4, -5).

1L-1,6-Di-O-benzyl-5-O-(2,3,4,6-tetra-O-benzyl- α -D-galactopyranosyl)-myo-inositol 8 and 1L-4,5-Di-O-benzyl-6-O-(2,3,4,6-tetra-O-benzyl- α -D-galactopyranosyl)-myo-inositol 9.—A mixture of compound 7 (0.44 g, 0.52 mmol), 4-methylmorpholine N-oxide monohydrate (NMO) (0.08 g, 0.62 mmol), and 0.05 mol dm⁻³ OsO₄ in toluene (1 cm³; 0.01 mmol) in acetone–water (8:1; 3 cm³) was stirred at room temperature for 16 h. After addition of NaHSO₃ (18 mg), the mixture was diluted with ethyl acetate (50 cm³), washed with brine, and dried. Evaporation of the solvent left a syrup, which was filtered on a bed of silica gel (5 g) to yield compounds 8 and 9 (0.42 g, 91%) as syrups in the ratio 2:1 (by ¹H NMR spectroscopy); [α]_D +7.6 (c 1.0, CHCl₃); δ_{H} (200 MHz; CDCl₃) 5.22 (1 H, d, J_{1,2}: 3.5, 1'-H), 3.15–5.18 (27 H, m, 1–6-H, 2'–6'-H, 3 × OH and 6 × CH₂Ph) and 7.15–7.45 (30 H, ArH).

1D-(1,3/2,4)-1-O-Acetyl-3,4-di-O-benzyl-2-O-(2,3,4,6-tetra-O-benzyl- α -D-galactopyranosyl)cyclohex-5-ene-1,2,3,4-tetraol 10.—Compound 7 (0.15 g, 0.17 mmol) was dissolved in pyridine (0.5 cm³), acetic anhydride (0.25 cm³) was added, and the solution was left at room temperature for 4 h. It was then diluted with water (100 cm³) and extracted into dichloromethane (100 cm³). The extract was washed successively with ice-cold 2% aq. sulfuric acid (20 cm³) and water, and dried. Evaporation of the solvent gave *title acetate 10* (0.18 g) as a syrup in quantitative yield (Found: C, 75.4; H, 6.5. C₅₆H₅₈O₁₀ requires C, 75.45; H, 6.59%); [α]_D +34 (c 1.0, CHCl₃); δ_{H} (200 MHz; CDCl₃) 1.91 (3 H, s, OAc), 3.45–4.98 (21 H, m, 1-, 5-, 6-H, 2'–6'-H, and 6 × CH₂Ph), 5.32–5.42 (1 H, m, 2-H), 5.44 (1 H, d, J_{1,2}: 3.4, 1'-H), 5.48 (1 H, AB-type quartet J_{3,4} 10, 4-H), 5.68 (1 H, AB-quartet, 3-H) and 7.05–7.35 (30 H, ArH).

1L-4-O-Acetyl-1,6-Di-O-benzyl-5-O-(2,3,4,6-tetra-O-benzyl- α -D-galactopyranosyl)-myo-inositol 11 and 1L-1-O-Acetyl-4,5-Di-O-benzyl-6-O-(2,3,4,6-tetra-O-benzyl- α -D-galactopyranosyl)-myo-inositol 12.—A mixture of compound 10 (0.19 g, 0.21 mmol), NMO (30 mg, 0.23 mmol) and 0.05 mol dm⁻³ OsO₄ in toluene (0.5 cm³, 0.005 mmol) in acetone–water (8:1; 2 cm³) was stirred at room temperature for 24 h. The usual work-up gave a syrup, which was filtered on a bed of silica gel (5 g) (eluted with 20% ethyl acetate in hexane) to yield *title compounds 11 and 12* (0.18 g, 92%) as syrups in the ratio 5.6:1; [α]_D +26.5 (c 1.0, CHCl₃); δ_{H} (200 MHz; CDCl₃) 2.04 (0.45 H, s, OAc), 2.07 (2.55 H, s, OAc), 3.40–5.15 (25.15 H, m, 1–6-H, 2'–6'-H, OH and 6 × CH₂Ph), 5.35 [0.85 H, t, J_{3,4} 8.0, 2(or 3)-H] and 5.52 (1 H, d, J_{1,2}: 3.2, 1'-H).

1L-4-O-Acetyl-2,3-di-O-benzoyl-1,6-di-O-benzyl-5-O-(2,3,4,6-tetra-O-benzyl- α -D-galactopyranosyl)-myo-inositol 13 and 1L-1-O-Acetyl-2,3-di-O-benzoyl-4,5-di-O-benzyl-6-O-(2,3,4,6-tetra-O-benzyl- α -D-galactopyranosyl)-myo-inositol 14.—To an ice-cold solution of compounds 11 and 12 (110 mg, 0.012 mmol) in pyridine (1 cm³) was added benzoyl chloride

(0.25 cm³) and the mixture was stirred at room temperature for 1 h before being poured into water, and extracted into dichloromethane. The extract was washed with 1% aq. hydrochloric acid (20 cm³), dried, and evaporated to obtain a syrup. This was filtered on silica gel (5 g) by elution with 5% ethyl acetate in hexane to obtain title compounds **13** and **14** as syrups in quantitative yield; [α]_D + 32.9 (c 1.0, CHCl₃); δ _H(200 MHz; CDCl₃) 1.92 (0.45 H, s, OAc), 1.96 (2.55 H, s, OAc), 3.50–4.90 (17 H, m, 1-, 5-, 6-H, 2'-6'-H and 4 × CH₂Ph), 5.18 (0.85 H, dd, $J_{4,3}$ 9.48, $J_{3,2}$ 2.6, 3-H), 5.41–5.50 (1 H, m, 4-H), 5.46 (0.15 H, dd, $J_{3,2}$ 2.89, $J_{3,4}$ 9.59, 3-H), 5.53 (1 H, d, $J_{1',2'}$ 3.3, 1'-H), 5.82 (0.85 H, t, $J_{3,2} = J_{1,2} = 2.68$, 2-H), 6.12 (0.15 H, t, $J_{1,2} = J_{2,3} = 2.72$, 2-H) and 7.05–8.35 (40 H, ArH).

References

- M. J. McConville, S. W. Homans, J. E. Thomas-Oates, A. Dell and A. Balic, *J. Biol. Chem.*, 1990, **265**, 7385; M. J. McConville, J. E. Thomas-Oates, M. A. J. Ferguson and S. W. Homans, *J. Biol. Chem.*, 1990, **265**, 19611; M. J. McConville and J. M. Blackwell, *J. Biol. Chem.*, 1991, **266**, 15170; P. M. Thomas and L. E. Samelson, *J. Biol. Chem.*, 1992, **267**, 12317.
- H. E. Carter, D. R. Strobach and J. N. Hawthorne, *Biochemistry*, 1969, **8**, 383; S. Steiner, S. Smith, C. J. Waechter and R. L. Lester, *Proc. Natl. Acad. Sci. USA*, 1969, **64**, 1042; S. W. Smith and R. L. Lester, *J. Biol. Chem.*, 1974, **249**, 3395; M. A. Ferguson, S. W. Homans, R. A. Dwek and T. W. Rademacher, *Science*, 1988, **239**, 753; S. W. Homans, C. J. Edge, M. A. Ferguson, R. A. Dwek and T. W. Rademacher, *Biochemistry*, 1989, **28**, 2881; B. Schmitz, R. A. Klien, I. A. Duncan, H. Egge, J. Gunawan, J. Peter-Katalinic, U. Dabrowski and J. Dabrowski, *Biochem. Biophys. Res. Commun.*, 1987, **146**, 1055; G. A. M. Cross, *Cell*, 1987, **48**, 179; M. G. Low, *Biochem. J.*, 1987, **244**, 1.
- C. E. Ballou and Y. C. Lee, *Biochemistry*, 1964, **3**, 682; Y. C. Lee and C. E. Ballou, *J. Biol. Chem.*, 1964, **239**, 1316.
- U. Malik and G. K. Kuller, *Indian J. Exp. Biol.*, 1983, **21**, 513.
- D. A. Cox, K. Richardson and B. C. Ross, *Topics in Antibiotic Chemistry*, ed. P. G. Sammes, Ellis Horwood, Sussex, 1977, vol. 1.
- W. Tanner and O. Kandler, *Plant Physiol.*, 1966, **41**, 1540; W. Tanner, L. Lehle and O. Kandler, *Biochem. Biophys. Res. Commun.*, 1967, **29**, 166; W. Tanner, *Z. Pflanzenphysiol.*, 1967, **57**, 474.
- D. R. Mootoo, P. Konradsson and B. Fraser-Reid, *J. Am. Chem. Soc.*, 1989, **111**, 8540.
- C. J. J. Elie, R. Verduyn, C. E. Dreef, G. A. van der Marel and J. H. van Boom, *J. Carbohydr. Chem.*, 1992, **11**, 715.
- S. Alenack, I. Kvarnstrom, A. Nikalsson, G. Nikalsson, S. C. T. Svensson and P. J. Garegg, *J. Carbohydr. Chem.*, 1991, **10**, 937.
- N. Chida, E. Yamada and S. Ogawa, *J. Carbohydr. Chem.*, 1988, **7**, 555.
- J. P. Vacca, S. J. de Solms, J. R. Hutt, D. C. Billington, R. Baker, J. J. Kulagowski and I. M. Mawer, *Tetrahedron*, 1989, **45**, 5679.
- A. E. Stepanov, B. A. Klyashchitskii, V. I. Shvets and R. P. Evstigneeva, *Bioorg. Khim.*, 1976, **2**, 1627.
- H. B. Meriyala and G. V. Reddy, *Tetrahedron*, 1991, **47**, 6435.
- S. Ken-ichi, S. Shogo, N. Yutaka, Y. Juji and H. Hironobu, *Chem. Lett.*, 1991, 17.
- R. J. Ferrier, *J. Chem. Soc., Perkin Trans. 1*, 1979, 1455; R. Blattner, R. J. Ferrier and S. R. Haines, *J. Chem. Soc., Perkin Trans. 1*, 1985, 2413; N. Chida, M. Ohtsuka, K. Nakazawa and S. Ogawa, *J. Org. Chem.*, 1991, **56**, 2976.
- S. Didier, P. Michel, D. Jeanne Marie, S. Anne-Marie and D. G. Stephan, *Synthesis*, 1983, 710.
- A. L. Gemal and J. L. Luche, *J. Am. Chem. Soc.*, 1981, **103**, 5454.
- H. A. J. Carless and K. Busia, *Tetrahedron Lett.*, 1990, **31**, 3449; S. V. Ley, *Pure Appl. Chem.*, 1990, **62**, 2031; D. A. Evans and S. W. Kaldor, *J. Org. Chem.*, 1990, **55**, 1698; J. K. Cha, W. J. Christl and Y. Kishi, *Tetrahedron*, 1984, **40**, 2247.
- V. A. Estevez and G. D. Bestwich, *Tetrahedron Lett.*, 1991, **32**, 623; C. Schultz, T. Metschies, B. Gerlach, C. Stadler and B. Jastroff, *Synlett.*, 1990, 1623.

Paper 2/05810C

Received 30th October 1992

Accepted 4th January 1993